DOI: 10.3779/j.issn.1009-3419.2018.09.03

### · Perspective ·

## *Helicobacter Pylori* Infection and Lung Cancer: New Insights and Future Challenges

Ileana GONZÁLEZ, Paulina ARAYA, Armando ROJAS

Biomedical Research Laboratories, Medicine Faculty, Catholic University of Maule, Talca, Chile

#### Abstract

*Helicobacter pylori* (*H. pylori*) is the causative agent of chronic gastritis and peptic ulcer diseases and is an important risk factor for the development functional dyspepsia, peptic ulceration, gastric adenocarcinoma and mucosa-associated lymphoid tissue lymphoma. *H. pylori* has very high rates of infection in human populations, and it is estimated that over 50% of the world population is infected. Recently, certain extra-gastric manifestations, linked to *H. pylori* infection, have been widely investigated. Noteworthy, a growing body of evidences supports an association between *H. pylori* infection with lung cancer. The present review intend to highlight not only the most recent evidences supporting this association, but also some missed points, which must be considered to validate this emerging association.

**Key words** *Helicobacter pylori;* Lung neoplasms; Air pollution; Pathogen-associated molecular patterns; Damage-associated molecular patterns; Chronic inflammation

#### Helicobacter pylori infection

Helicobacter pylori (H. pylori) is a Gram-negative spiralshaped bacterium that persistently colonizes the human stomach. In this sense, colonization constitutes an established and major risk factor in the pathogenesis of functional dyspepsia, peptic ulceration, gastric adenocarcinoma and mucosa-associated lymphoid tissue lymphoma<sup>[1]</sup>. In fact, and since 1994 H. pylori has been classified as a Group 1 carcinogen the International Agency for Research on Cancer (IARC)<sup>[2]</sup>.

*H. pylori* chronically infects more than half of the world's population, being estimated that this gramnegative bacterium has co-evolved with its human host since its migration out of Africa along with human host approximately 60,000 years ago<sup>[3]</sup>.

The pathogenesis of *H. pylori* mainly depends on the exposure of several bacterial factors to the host, including cytotoxin-associated gene A (CagA), vacuolating cytotoxin A (VacA), type IV secretion system (T4SS), outer inflammatory protein A, as well as different adherence factors. Because of their critical roles in *H. pylori*-induced pathogenesis, these pathogenicity factors have been extensively studied not only in gastric cancer high-risk *H. pylori*-infected populations but also on their carcinogenic mechanisms. Although the

vast majority of *H. pylori* in colonized hosts are free-living, and approximately 20% bind to gastric epithelial cells and adherence is required for prolonged persistence in the stomach and for induction of injury<sup>[4-6]</sup>.

More recently, the scope of the pathogenic mechanisms linked to the oncogenic potential of *H. pylori* has been extended to the capacity of this pathogen to produce genomic instability on gastric epithelial cells<sup>[7]</sup>.

Strikingly, gene products from the *cag* pathogenic island (PAI) appear to play an important role in the accumulation of DNA double-strand breaks (DSBs) in infected host cells and the expression of RAD51 is reduced after infection with *cag*-positive strains<sup>[8]</sup>.

Indeed, RAD51 plays an important role in homologous strand exchange, a key step in DNA repair of double strand breaks through homologous recombination (HR)<sup>[9]</sup>.

Additionally, the interactions of *H. pylori* with host cells result initially to the adhesion to epithelium, which in turn induce a marked inflammatory responses resulting in persistent colonization, chronic inflammation and severe inflammation, the disruption of the epithelial barrier function<sup>[10-12]</sup>.

Furthermore, the *H. pylori* associated gastric cancer is characterized by a chronic inflammatory phenotype, where the long-lasting activation of the key inflammatory regulator nuclear factor kappa B (NF- $\kappa$ B) is essential in contributing to neoplastic transformation<sup>[13,14]</sup>. At the cellular level, myeloid and lymphocytic cells frequently infiltrate malignant lesions.



Correspondence to: Ileana GONZALEZ, E-mail: ileanag@ucm.cl; Armando ROJAS, E-mail: arojasr@ucm.cl

Tumor-associated macrophages (TAM) promote malignant progression and the degree of TAMs infiltration correlates with tumor progression and clinical disease stage<sup>[15]</sup>.

#### Extra-digestive diseases and H. pylori infection

During the late 1990s the first reports showing that *H*. *pylori* is associated with extra-digestive diseases appeared in scientific literature<sup>[16-20]</sup>.

Since then, a huge body of evidences has support the association of *H. pylori* infection with many extra-digestive diseases including autoimmune diseases, cardiovascular diseases, colonic and pancreatic diseases, diabetes mellitus, hepatobiliary system diseases, neurological disorders, skin diseases, hematological diseases and as well as reproductive disorders<sup>[21-23]</sup>.

The immune and inflammatory responses triggered by *H. pylori* infection is postulated as the main mechanisms supporting the extra-digestive pathologies<sup>[24]</sup>.

Of note, recent studies of seroprevalence of *H. pylori* in patients, as well as some *meta*-analysis studies have also suggested a significant association between *H. pylori* infection and chronic respiratory diseases<sup>[25-29]</sup>. However the pathogenetic mechanism(s) for the effects of *H. pylori* in lungs remains mainly elusive.

#### The case of lung cancer

Lung cancer remains not only as the leading type of cancer worldwide, but also as as the most common cause of death from cancer<sup>[30]</sup>. Primary lung cancer is known to be caused by either voluntary or involuntary inhalation of environmental carcinogens and where lung cancer cases attributable to smoking has reached up to 90% in countries with a history of tobacco consumption, as reported by IARC<sup>[31]</sup>.

However, 10%-15% of lung cancer cases diagnosed in Western countries and approximately one of every four cases in Asia occur in never smoker subjects<sup>[32]</sup>.

Infectious agents and their potential roles in carcinogenesis have been extensively investigated for many years. In fact, it is estimated that almost 25% of all cancers are somehow associated with chronic infection and inflammation.

Accordingly, several evidences derived from both, epidemiological studies and basic research, have shown that organ-specific carcinogenesis is linked to the development of a chronic and local inflammatory milieu raised as a consequence of host response to infection, as demonstrated for *H. pylori* infection and gastric cancer, *Salmonella typhi* and gallbladder carcinoma, human papilloma virus with both oropharyngeal and cervical cancers, and nasopharyngeal carcinoma and Hodgkin's lymphoma with Epstein-Bar virus, just to mention a few<sup>[33-37]</sup>.

In this context, and on spite that some published studies are controversial, the most recent reports support the association between *H. pylori* infection and lung cancer<sup>[38]</sup>.

Noteworthy, some recent studies have shed light onto the pathogenic mechanisms involved in this association. Very interestingly, the presence of DNA from *H. pylori* detected by Real-time PCR and even some pathogen-derived proteins such as Vac-A, have been found in bronchoalveolar lavage from lung cancer and in lung biopsies specimens, respectively. Additionally, VacA is able to induce induces both IL-6 and IL-8 production in the lung carcinoma cell line A549, as well as Il-8 in human bronchial epithelial cells, just supporting the idea de lung epithelium is responsive to pathogenic factors of *H. pylori*<sup>[39,40]</sup>.

To understand how these bacterial components can be found on lung tissue it is important to highlight that available clinical and experimental evidence points to a possible relationship between the progression of airways disease, pro-inflammatory processes and gastric aspiration<sup>[41,42]</sup>, and where the main cause is thought to be due tracheobronchial aspiration of small amounts of stomach-associated components, such as pepsin or bile acids, and thus causing repetitive subclinical injury to the lung<sup>[43,44]</sup>.

Additionally, oral cavity has been suggested as an extragastric reservoir of *H. pylori*, and thus this pathogen can reach the lungs from either stomach or oral cavity<sup>[45-47]</sup>.

In the case that either *H. pylori* or some of its components reach the pulmonary epithelium, it would trigger without any doubt an inflammatory response.

The epithelial lining of the lung abundantly expresses pathogen-recognition receptors (PRRs), like Toll-like receptors (TLRs) to detect a myriad of pathogen-associated molecular patterns (PAMPS). Either lung epithelial cells, alveolar macrophages or dendritic cells, which constitute the first line of lung defense, express TLRs on their surfaces able to recognize not only bacterial-associated PAMPS but also cell-wall and membrane components such as peptidoglycan, lipoproteins, lipoteichoic acid or even pathogen-secreted toxins<sup>[48]</sup>. Furhermore pathogen DNA is recognized by cytoplasmatic surveillance receptors such as TLR-9.

In this context, the receptor of advanced glycation end-products (RAGE) is now recognized as a pathogenrecognition receptor<sup>[49]</sup>. This receptor is abundantly expressed at type-I alveolar epithelial cells (AT1), which comprise only 4% of the alveolar surface area, yet they constitute 60% of alveolar epithelial cells and 10%-15% of all lung cells. AT1 cells are large squamous cells that cover 95% of the alveolar surface area and form the epithelial

中国肺癌杂志 www.lungca.org component of the thin air-blood barrier<sup>[50]</sup>. Additionally, RAGE is also expressed in differentiating alveolar type-II epithelial cells (AT2), bronchial smooth muscle cells, vascular endothelial cells, and pulmonary macrophages<sup>[51]</sup>.

Noteworthy, TLR-2, TLR-4 and RAGE are involved in the recognition of PAMPS in *H. pylori* and thus triggering robust inflammatory response, not only at gastric epithelium<sup>[52]</sup> but also in monocytes/macrophages<sup>[53]</sup>, dendritic cells<sup>[54]</sup> and B cells<sup>[55]</sup>.

Interestingly, both TLRs and RAGE can response either directly, by recognizing PAMPs, but also indirectly through the recognition of damage-associated molecular patterns (DAMPs), also known as alarmins, generated as a consequence of cellular stress, damage or cell death, and where the release of DAMPs as a consequence of lung injury has been extensively reported<sup>[56-60]</sup>.

Furhermore, epithelial cell-derived expression of inflammatory mediators after of PAMPs or DAMPs recognition by PPRs, markedly influences the recruitment and activation of immune cells responses in the lungs<sup>[61]</sup>.

Therefore, the chronic exposure to pathogens or pathogens-derived components, oxidants and toxic pollutants causes the release of DAMPs that activate epithelial cellintrinsic pattern-recognition pathways and also recruit and activate cells of the immune system.

The contribution of air of pollution/smoking deserves a special attention. Of note, it is estimated that there are nearly 1 billion smokers worldwide and approximately 80% live in either low- or middle-income countries where the effects or burdens of tobacco-related illness and death, such as lung cancer, are the most documented<sup>[62]</sup>.

In addition, the World Health Organization (WHO) has estimated that 91% of the world's population lives in places where air quality levels exceed WHO limits, being the lowand middle-income countries those that experience the highest burden<sup>[63]</sup>. On the other hand, about 50% of the world population is infected by *H. pylori* and the rate of infections varies from 15.5% for high-incomes countries up to 93.6% for in low- and middle-incomes countries<sup>[64]</sup>.

Therefore, it is likely that a high number of *H. pylori*infected subjects living in low and middle-incomes countries are also exposed to high levels of air pollutants. In this context, it is tempting to speculate, that chronic and subclinical tracheobronchial aspiration in *H. pylori*infected subjects, together with the burden of smoking or air-pollution can act synergically to establish and perpetuate an inflammatory reaction at epithelial lining of the lung, and thus favoring malignant transformation and tumor growth.

Based on the most recent reports suggesting the association between *H. pylori* infection and chronic respiratory diseases, and particularly lung cancer, further studies are imperative not only to validate the association, but also to understand the contribution of other factors such as smoking habits or air pollution, and where the underlying molecular mechanisms still remains to be clarified.

#### References

- Cavaleiro-Pinto M, Peleteiro B, Lunet N, et al. Helicobacter pylori infection and gastric cardia cancer: systematic review and metaanalysis. Cancer Causes Control, 2011, 22(3): 375-387. doi: 10.1007/ s10552-010-9707-2
- Schistosomes, liver flukes and Helicobacter pylori. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Lyon, 7-14 June 1994. IARC Monogr Eval Carcinog Risks Hum 1994; 61: 1-241.
- 3 Correa P, Piazuelo MB. Evolutionary history of the Helicobacter pylori genome: Implications for gastric carcinogenesis. Gut Liver, 2012, 6(1): 21-28. doi: 10.5009/gnl.2012.6.1.21
- Kabamba ET, Tuan VP, Yamaoka Y. Genetic populations and virulence factors of Helicobacter pylori. Infect Genet Evol, 2018(4), 60: 109-116. doi: 10.1016/j.meegid.2018.02.022
- 5 Nejati S, Karkhah A, Darvish H, et al. Influence of Helicobacter pylori virulence factors CagA and VacA on pathogenesis of gastrointestinal disorders. Microb Pathog, 2018, 117: 43-48. doi: 10.1016/j.micpath.2018.02.016
- 6 Camilo V, Sugiyama T, Touati E. Pathogenesis of *Helicobacter pylori* infection. Helicobacter, 2017, 22(Suppl 1): e12405. doi: 10.1111/ hel.12405
- 7 Toller IM, Neelsen KJ, Steger M, et al. Carcinogenic bacterial pathogen Helicobacter pylori triggers DNA double-strand breaks and a DNA damage response in its host cells. Proc Natl Acad Sci U S A, 2011, 108(36): 14944-14949. doi: 10.1073/pnas.1100959108
- 8 Hanada K, Uchida T, Tsukamoto Y, et al. Helicobacter pylori infection introduces DNA double-strand breaks in host cells. Infect Immun, 2014, 82(10): 4182-4189. doi: 10.1128/IAI.02368-14
- 9 Tarsounas M, Davies A, West S. RAD51 localization and activation following DNA damage. Philos Trans R Soc Lond B Biol Sci, 2004, 359(1441): 87-93. doi: 10.1098/rstb.2003.1368
- 10 Suzuki M, Mori M, Miyayama A, *et al.* Enhancement of neutrophil infiltration in the corpus after failure of Helicobacter pylori eradication. J Clin Gastroenterol, 1997, 25 Suppl 1: S222-S228.
- Bruewer M, Luegering A, Kucharzik T, et al. Proinflammatory cytokines disrupt epithelial barrier function by apoptosisindependent mechanisms. J Immunol, 2003, 171(11): 6164-6172. doi: 10.4049/jimmunol.171.11.6164.
- 12 Tran LS, Chonwerawong M, Ferrero RL. Regulation and functions of inflammasome-mediated cytokines in Helicobacter pylori infection. Microbes Infect, 2017, 19(9-10): 449-458. doi: 10.1016/ j.micinf.2017.06.005.
- 13 Sokolova O, Borgmann M, Rieke C, *et al*. Helicobacter pylori induces type 4 secretion system-dependent, but CagA-independent activation

# 中国肺癌杂志 www.lungca.org

#### 中国肺癌杂志2018年9月第21卷第9期 Chin J Lung Cancer, September 2018, Vol.21, No.9

of IkappaBs and NF-kappaB/RelA at early time points. J Immunol, 2003, 171(11): 6164-6172. doi:10.1016/j.ijmm.2013.07.008.

- Schweitzer K, Sokolova O, Bozko PM, et al. Helicobacter pylori induces NF-kappaB independent of CagA. EMBO Rep, 2010, 11(1): 10-11. doi: 10.1038/embor.2009.263
- 15 Rojas A, Delgado-López F, Gonzalez I. Tumor-associated macrophages in gastric cancer: more than bystanders in tumor microenvironment. Gastric Cancer, 2017, 20(1): 215-216. doi: 10.1007/s10120-016-0596-2.
- 16 Perez-Perez GI, Peek RM, Legath AJ, et al. The role of CagA status in gastric and extragastric complications of Helicobacter pylori. J Physiol Pharmacol, 1999, 50(5): 833-845.
- Moran AP. Helicobacter pylori lipopolysaccharide-mediated gastric and extragastric pathology. J Physiol Pharmacol, 1999, 50(5): 787-805.
- 18 Konturek SJ, Konturek PC, Pieniazek P, et al. Role of Helicobacter pylori infection in extragastroduodenal disorders: introductory remarks. J Physiol Pharmacol, 1999, 50(5): 683-694.
- 19 Gasbarrini A, Franceschi F. Autoimmune diseases and Helicobacter pylori infection. Biomed Pharmacother, 1999, 53(5-6): 223-226.
- 20 Konstacký M. Relation between Helicobacter pylori infection and ischemic heart disease. Cas Lek Cesk, 1998, 137(16): 483-485.
- 21 de Korwin JD, Ianiro G, Gibiino G, et al. Helicobacter pylori infection and extragastric diseases in 2017. Helicobacter, 2017, 22 Suppl 1: 123. doi:10.1111/hel.12411
- 22 Chmiela M, Gonciarz W. Molecular mimicry in Helicobacter pylori infections. World J Gastroenterol, 2017, 23(22): 3964-3977. doi: 10.3748/wjg.v23.i22.3964
- Kyburz A, Müller A. Helicobacter pylori and extragastric diseases. Curr Top Microbiol Immunol, 2017, 400: 325-347. Doi: 10.1007/97 8-3-319-50520-6
- 24 Salama NR, Hartung ML, Müller A. Life in the human stomach: persistence strategies of the bacterial pathogen Helicobacter pylori. Nat Rev Microbiol, 2013, 11(6): 385-399. doi: 10.1038/nrmicro3016
- 25 Tsang KW, Lam SK, Lam WK, et al. High seroprevalence of Helicobacter pylori in active bronchiectasis. Am J Respir Crit Care Med, 1998, 158(4): 1047-1051.
- 26 M. Caselli, E. Zaffoni, M. Ruina, *et al.* Helicobacter pylori and chronic bronchitis. Scand J Gastroenterol, 1999, 34(8): 828-830.
- 27 Bennett D, Bargagli E, Refini RM, et al. Helicobacter pylori seroprevalence in patients with idiopathic pulmonary fibrosis. Eur Respir J, 2014, 43(2): 635-638. doi: 10.1183/09031936.00104813
- Wang L, Guan Y, Li Y, et al. Association between chronic respiratory diseases and Helicobacter pylori: A meta-analysis. Arch Bronconeumol, 2015, 51(6): 273-278. doi: 10.1016/j.arbres.2014.03.019
- 29 Wang F, Liu J, Zhang Y, et al. Association of Helicobacter pylori infection with chronic obstructive pulmonary disease and chronic bronchitis: a meta-analysis of 16 studies. Infect Dis (Lond), 2015, 47(9): 597-603. doi: 10.3109/00365548.2014.989539
- 30 Ferlay J , Soerjomataram I, Dikshit R, et al. Cancer incidence and

mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer, 2015, 136(5): E359-E386. doi: 10.1002/ijc.29210

- 31 IARC, Tobacco Smoke and Involuntary Smoking. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, Vol. 83, International Association for Research on Cancer, Lyon, 2004.
- 32 Sun S, Schiller J, Gazdar A. Lung cancer in never smokers--a different disease. Nat Rev Cancer, 2007, 7(10): 778-790. doi: 10.1038/nrc2190
- 33 Pormohammad A, Ghotaslou R, Leylabadlo HE, et al. Risk of gastric cancer in association with Helicobacter pylori different virulence factors: A systematic review and meta-analysis. Microb Pathog, 2018, 118: 214-219. doi: 10.1016/j.micpath.2018.03.004
- 34 Scanu T, Spaapen RM, Bakker JM, et al. Salmonella manipulation of host signaling pathways provokes cellular transformation associated with gallbladder carcinoma. Cell Host Microbe, 2015, 17(6): 763-774. doi: 10.1016/j.chom.2015.05.002
- 35 Yete S, D'Souza W, Saranath D. High-risk human papillomavirus in oral cancer: clinical implications. Oncology, 2018, 94(3): 133-141. doi: 10.1159/000485322.
- 36 Oyervide M, Pérez A, Rodríguez H, et al. Understanding the HPV integration and its progression to cervical cancer. Infect Genet Evol, 2018, 61: 134-144. doi: 10.1016/j.meegid.2018.03.003
- 37 Dolcetti R. Cross-talk between Epstein-Barr virus and microenvironment in the pathogenesis of lymphomas. Semin Cancer Biol, 2015, 34: 58-69. doi: 10.1016/j.semcancer.2015.04.006.
- 38 Mounika P. Helicobacter pylori infection and risk of lung cancer: A meta-analysis. Lung Cancer Int, 2013, 2013: 131869. doi: 10.1155/2013/131869.
- 39 Samareh-Fekri M, Hashemi Bajgani SM, Shafahi A, et al. Detection of Helicobacter pylori in the bronchoalveolar lavage of patients with lung cancer using real-time PCR. Jundishapur J Microbiol, 2016, 9(11): e32144. doi: 10.5812/jjm.32144.
- 40 Nakashima S, Kakugawa T, Yura H, et al. Identification of Helicobacter pylori VacA in human lung and its effects on lung cells. Biochem Biophys Res Commun, 2015, 460(3): 721-726. doi: 10.1016/j.bbrc.2015.03.096.
- 41 Hunt E, Sullivan A, Galvin J, et al. Gastric aspiration and Its role in airway inflammation. Open Respir Med J, 2018, 12: 1-10. doi: 10.2174/1874306401812010001.
- 42 Zerbib F, Dulery C. Facts and fantasies on extraesophageal reflux: A gastroenterologist perspective. J Clin Gastroenterol, 2017, 51(9): 769-776. doi: 10.1097/MCG.000000000000918.
- Lee J, Collard H, Raghu G, *et al*. Does chronic microaspiration cause idiopathic pulmonary fibrosis? Am J Med, 2010, 123(4): 304-311. doi: 10.1016/j.amjmed.2009.07.033.
- 44 Starosta V, Kitz R, Hartl D, *et al.* Bronchoalveolar pepsin, bile acids, oxidation, and inflammation in children with gastroesophageal reflux disease. Chest, 2007, 132(5): 1557-1564. doi: 10.1378/ chest.07-0316.
- 45 Payão S, Rasmussen L. Helicobacter pylori and its reservoirs:

中国肺癌杂志 www.lungca.org 中国肺癌杂志2018年9月第21卷第9期 Chin J Lung Cancer, September 2018, Vol.21, No.9

A correlation with the gastric infection. World J Gastrointest Pharmacol Ther, 2016, 7(1): 126-132. doi: 10.4292/wjgpt.v7.i1.126.

- Al Sayed A, Anand P, Kamath K, *et al.* Oral cavity as an extragastric reservoir of Helicobacter pylori. ISRN Gastroenterol, 2014: 261369.
  doi: 10.1155/2014/261369.
- Assumpção M, Martins L, Melo Barbosa H, et al. Helicobacter pylori in dental plaque and stomach of patients from Northern Brazil. World J Gastroenterol, 2010, 16(24): 3033-3039. doi: 10.3748/wjg. v16.i24.3033.
- Kawai T, Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. Immunity, 2011, 34(5): 637-650. doi: 10.1016/j.immuni.2011.05.006.
- 49 Rojas A, Pérez-Castro R, González I, *et al.* The emerging role of the receptor for advanced glycation end products on innate immunity. Int Rev Immunol, 2014, 33(1): 67-80. doi: 10.3109/08830185.2013. 849702.
- 50 Yang J, Hernandez B, Martinez-Alanis D, et al. The development and plasticity of alveolar type 1 cells. Development, 2016, 143(1): 54-65. doi: 10.1242/dev.130005.
- 51 Downs C, Kreiner L, Johnson N, et al. Receptor for advanced glycation end-products regulates lung fluid balance via protein kinase C-gp91 (phox) signaling to epithelial sodium channels. Am J Respir Cell Mol Biol, 2015, 52(1): 75-87. doi: 10.1165/rcmb.2014-0002OC.
- 52 Rojas A, González I, Rodríguez B, et al. Evidence of involvement of the receptor for advanced glycation end-products (RAGE) in the adhesion of Helicobacter pylori to gastric epithelial cells. Microbes Infect, 2011, 13(10): 818-823. doi: 10.1016/j.micinf.2011.04.005.
- 53 Obonyo M, Sabet M, Cole SP, et al. Deficiencies of myeloid differentiation factor 88, Toll-like receptor 2 (TLR2), or TLR4 produce specific defects in macrophage cytokine secretion induced by Helicobacter pylori. Infect Immun, 2007, 75(5): 2408-2414. doi: 10.1128/IAI.01794-06.
- 54 Kim DJ, Park JH, Franchi L, et al. The Cag pathogenicity island and interaction between TLR2/NOD2 and NLRP3 regulate IL-1β production in Helicobacter pylori infected dendritic cells. Eur J Immunol, 2013, 43(10): 2650-2658. doi: 10.1002/eji.201243281.

- 55 Sayi A, Kohler E, Toller IM, et al. TLR-2-activated B cells suppress Helicobacter-induced preneoplastic gastric immunopathology by inducing T regulatory-1 cells. J Immunol, 2011, 186(2): 878-890. doi: 10.4049/jimmunol.1002269.
- 56 Lagiedo M, Sikora J, Kaczmarek M. Damage associated molecular patterns in the course of lung cancer-A review. Scand J Immunol, 2015, 82(2): 95-101. doi: 10.1111/sji.12308.
- Fujishima S. Pathophysiology and biomarkers of acute respiratory distress syndrome. J Intensive Care, 2014, 2(1): 32. doi: 10.1186/2052-0492-2-32.
- 58 Xiang M, Fan J. Pattern recognition receptor-dependent mechanisms of acute lung injury. Mol Med, 2010, 16(1-2): 69-82. doi: 10.2119/ molmed.2009.00097.
- 59 Kang JH, Hwang SM, Chung IY. S100A8, S100A9, and S100A12 activate airway epithelial cells to produce MUC5AC via ERK and NF-κB pathways. Immunology, 2015, 144(1): 79-90. doi: 10.1111/ imm.12352.
- 60 Ellson C, Dunmore R, Hogaboam C, et al. Danger-associated molecular patterns and danger signals in idiopathic pulmonary fibrosis. Am J Respir Cell Mol Biol, 2014, 51(2): 163-168. doi: 10.1165/rcmb.2013-0366TR.
- 61 Whitsett JA, Alenghat T. Respiratory epithelial cells orchestrate pulmonary innate immunity. Nat Immunol, 2015, 16(1): 27-35. doi: 10.1038/ni.3045.
- 62 Mackay JE, Shafey O. The Tobacco Atlas. 2nd ed. American Cancer Society; Atlanta, GA, USA: 2006.
- 63 World Health Organization. Ambient air pollution: a global assessment of exposure and burden of disease. World Health Organization 2016. http://www.who.int/iris/handle/10665/250141.
- 64 Eusebi LH, Zagari RM, Bazzoli F. 2014. Epidemiology of Helicobacter pylori infection. Helicobacter, 2014, 19(1): 1-5. doi: 10.1111/hel.12165.

(Received: 2018-05-10 Revised: 2018-06-20 Accepted: 2018-06-25) (Edited by Juan NAN)



Cite this article as: Gonzalez I, Araya P, Rojas A. *Helicobacter Pylori* Infection and Lung Cancer: New Insights and Future Challenges. Zhongguo Fei Ai Za Zhi, 2018, 21(9): 658-662. doi: 10.3779/j.issn.1009-3419.2018.09.03



• 662 •